

Amplite™ Fluorimetric NADP/NADPH Assay Kit

Red Fluorescence

Ordering Information:

Product Number: #15259 (400 assays)

Instrument Platform:

Fluorescence microplate readers

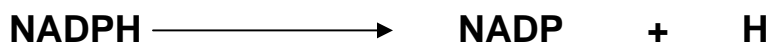
Storage Conditions:

Keep in freezer and avoid light

Introduction

Nicotinamide adenine dinucleotide (NAD⁺) and nicotinamide adenine dinucleotide phosphate (NADP⁺) are two important cofactors found in cells. NADH is the reduced form of NAD⁺, and NAD⁺ is the oxidized form of NADH. It forms NADP with the addition of a phosphate group to the 2' position of the adenyl nucleotide through an ester linkage. NADP is used in anabolic biological reactions, such as fatty acid and nucleic acid synthesis, which require NADPH as a reducing agent. In chloroplasts, NADP is an oxidizing agent important in the preliminary reactions of photosynthesis. The NADPH produced by photosynthesis is used as reducing power for the biosynthetic reactions in the Calvin cycle of photosynthesis. The traditional NAD/NADH and NADP/NADPH assays are done by monitoring of NADH or NADPH absorption at 340 nm. This method suffers low sensitivity and high interference since the assay is done in the UV range that requires expensive quartz microplate.

This Amplite™ NADP/NADPH Assay Kit provides a convenient method for sensitive detection of NADP, NADPH and their ratio. The enzymes in the system specifically recognize NADP/NADPH in an enzyme cycling reaction. There is no need to purify NADP/NADPH from sample mix. The enzyme cycling reaction significantly increases detection sensitivity. In addition, this assay has very low background since it is run in the red visible range that significantly reduces the interference from biological samples. The assay has demonstrated high sensitivity and low interference with 570 nm excitation 590 nm emission.



NADP Formation
is monitored by
NADPH Sensor

The Amplite™ Fluorimetric NADP/NADPH Assay Kit provides a sensitive, one-step fluorimetric assay to detect as little as 1 picomoles of NADP (H) in a 100 µL assay volume (10 nM; Figure 1). The assay can be performed in a convenient 96-well or 384-well microtiter-plate format and easily adapted to automation with no separation steps required. Its signal can be easily read by either fluorescence microplate reader with

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Ex/Em = 530 to 570 nm/590 to 600 nm (maximum Ex/Em = 570 nm/590 nm) or absorbance microplate reader at 576±5 nm.

Kit Key Features

Broad Application: Can be used for quantifying NADP/NADPH in solutions, in cell extracts.

Sensitive: The kit detect as low as 1 picomoles of NADP/NADPH in solution.

Continuous: Easily adapted to automation with no separation required.

Convenient: Formulated to have minimal hands-on time. No wash is required.

Non-Radioactive: No special requirements for waste treatment.

Kit Components

Component	Amount
Component A: NADP/NADPH recycling enzyme mixture	2 bottles (lyophilized powder)
Component B: NADPH sensor buffer	1 bottle (20 mL)
Component C: NADPH standard (FW: 833.36)	1 vial (167 µg)

Assay Protocol for one 96-well plate

Brief Summary

Prepare NADP/NADPH reaction mixture (50 µL) → Add NADPH standards or test samples (50 µL)
→ Incubate at room temperature for 15 min-2hr → Read fluorescence at Ex 570 nm/Em 590

Note: Thaw 1 vial (or bottle) each of all the kit components to room temperature before starting your experiment.

1. Prepare NADPH stock solution:

Prepare NADPH standard stock solution: Add 200 µL of PBS buffer into the NADPH standard vial (Component C) to make 1 mM (1 nmol/µL) stock solution.

Note: The unused NADPH solution should be divided as single use aliquots and stored at -20°C.

2. Prepare NADP/NADPH reaction mixture:

Prepare the NADP/NADPH reaction mixture: Add 10 mL of NADP/NADPH sensor buffer (Component B) to the bottle of NADP/NADPH recycling enzyme mixture (Component A), mixed well.

Note: This solution is enough for two 96-well plates. The unused NADP/NADPH mixture should be divided as single use aliquots and stored at -20°C.

3. Prepare Serial NADPH (0 to 10 µM) solutions

3.1 Add 10 µL of NADPH standard stock solution (from step 1) to 990 µL PBS buffer to generate 10 µM (10 pmol/µL) standard.

Note: Diluted NADPH standard solution is unstable, should be used within 4 hours.

- 3.2 Take 200 μL of 10 μM solution to perform 1:3 serial dilutions to get 3, 1, 0.3, 0.1, 0.03, 0.01, 0.03 and 0 standard NADPH solutions.
- 3.3 Add NADPH standards and NADP/NADPH-containing test samples into a 96-well solid black microplate as described in Tables 1 and 2

Note: Prepare your cell or tissue samples as desired.

Table 1. Layout of NADPH standards and test samples in a solid black 96-well microplate:

BL	BL	TS	TS						
NS1	NS1						
NS2	NS2										
NS3	NS3										
NS4	NS4										
NS5	NS5										
NS6	NS6										
NS7	NS7										

Note: NS= NADPH standards, BL=Blank control, TS=test samples.

Table 2. Reagent composition for each well:

NADPH Standard	Blank Control	Test Sample
Serial dilutions* (50 μL)	PBS: 50 μL	50 μL

**Note: Add the serially diluted NADPH standards from 0.003 μM to 3 μM into wells from NS1 to NS7 in duplicate.*

High concentration of NADPH (e.g., >100 μM , final concentration) may cause reduced fluorescence signal due to the overoxidation of NADPH sensor (to a non-fluorescent product).

4. Run NADP/NADPH Assay in supernatants

- 4.1 Add 50 μL of NADPH reaction mixture (from step 2) to each well of the NADPH standard, blank control, and test samples (see step 3.3) so that the total NADPH assay volume is 100 μL /well
Note: For a 384-well plate, add 25 μL sample, 25 μL of NADPH reaction mixture per well.

- 4.2 Incubate the reaction for 15 minutes to 2 hours at room temperature, protected from light.

- 4.3 Monitor the fluorescence increase with 530-570 nm (optimal at 570) excitation and 590-600 nm emission by using a fluorescence plate reader.

Note1: The contents of the plate can also be transferred to a white clear bottom plate and read by absorbance microplate reader at the wavelength of 576 ± 5 nm. The absorption detection has lower sensitivity compared to fluorescence reading.

Note2: To detect NADPH only, aliquot 200 μL samples into Eppendorf tubes. Heat samples to 60°C for 30 min in a heating block or a water bath. All NADP will be destroyed while NADPH will be still intact under these conditions. Cool samples on ice, and quickly spin samples if precipitates occur. Transfer 50 μL of NADPH samples into the wells as indicated in Table 1 and 2.

5. Run Data Analysis

The fluorescence in blank wells (with the PBS buffer only) is used as a control, and is subtracted from the values for the wells of NADPH reactions. The typical data are shown in Figure 1 (NADPH standard curve).

Note: The fluorescence background increases with time, thus it is important to subtract the fluorescence intensity value of the blank wells for each data point.

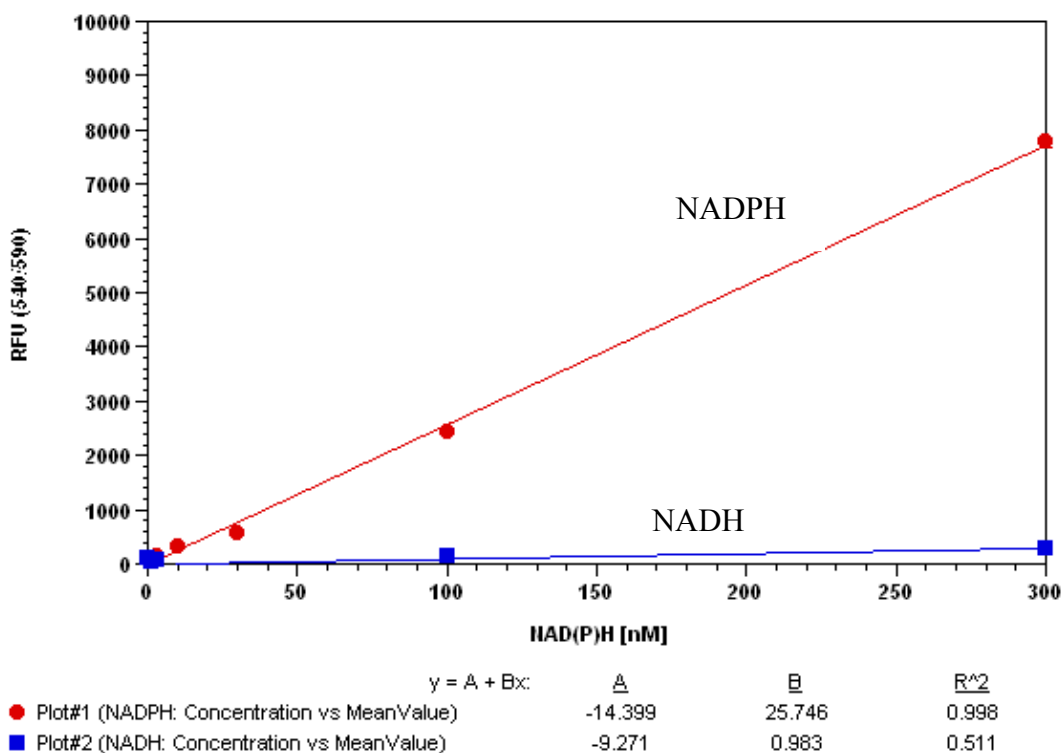


Figure 1. NADPH dose response on 96-well black plate was measured with Amplite™ NADP/NADPH Assay Kit using a BMG LabTech NOVOSTar microplate reader. As low as 10 nM (1 pmol/well) of NADPH can be detected with 30 min incubation time (n=3) while there is no response from NADH.

Related Products

15257	Amplite™ Fluorimetric NAD/NADH Assay Kit *Red Fluorescence*	1 kit
15258	Amplite™ Colorimetric NAD/NADH Assay Kit	1 kit
15259	Amplite™ Fluorimetric NADP/NADPH Assay Kit *Red Fluorescence*	1 kit
15260	Amplite™ Colorimetric NADP/NADPH Assay Kit	1 kit
15261	Amplite™ Fluorimetric NADH Assay Kit *Red Fluorescence*	1 kit
15262	Amplite™ Fluorimetric NADPH Assay Kit *Red Fluorescence*	1 kit

References:

1. Hedeskov CJ, Capito K, Thams P. (1987) Cytosolic ratios of free [NADPH]/[NADP+] and [NADH]/[NAD+] in mouse pancreatic islets, and nutrient-induced insulin secretion. *Biochem J*, 241, 161.
2. Gaetani GF, Ferraris AM, Sanna P, Kirkman HN. (2005) A novel NADPH:(bound) NADP+ reductase and NADH:(bound) NADP+ transhydrogenase function in bovine liver catalase. *Biochem J*, 385, 763.
3. Kobayashi K, Miura S, Miki M, Ichikawa Y, Tagawa S. (1995) Interaction of NADPH-adrenodoxin reductase with NADP+ as studied by pulse radiolysis. *Biochemistry*, 34, 12932.
4. Marino D, Gonzalez EM, Frendo P, Puppo A, Arrese-Igor C. (2006) NADPH recycling systems in oxidative stressed pea nodules: a key role for the NADP(+)-dependent isocitrate dehydrogenase. *Planta*.