AmpliteTM Fluorimetric Thiol Quantitation Kit *Green Fluorescence*

Ordering Information: Product number: 5524 (200 assays)

Instrument Platform: Fluorescence microplate readers

Storage Conditions: Keep in -20°C avoid moisture and light

Introduction

The detection and measurement of free thiol (such as free cysteine, glutathione and cysteine residues in proteins) is one of the essential tasks for investigating biological processes and events in many biological systems. The thiol of cysteine is a powerful nucleophile, and cysteine is the most easily oxidized of the amino acids, forming covalent crosslinks between peptide chains. Because of the unique reactivity, cysteines in proteins have been the site, both deliberately and inadvertently, of a vast number of modification reactions. These modifications include oxidations with a variety of oxidizing agents, reaction with a-halo acids and organic mercurial derivatives, and the reversible formation of disulfides by reaction with a wide variety of reagents including tetrathionate to produce *S*-sulfocysteine and alkyl or aryl methanethiosulfonates to yield organic disulfides.

The monitoring of reduced (GSH) and oxidized glutathione in biological samples is essential for evaluating the redox and detoxification status of cells and tissues in relation to the protective role of glutathione against oxidative and free-radical-mediated cell injury. Disorders of cysteine metabolism include cystinosis, an autosomal recessive disease produced by a defect in lysosomal transport, and cystinuria, a common heritable disorder of amino acid transport. Cysteine is unique among the amino acids found in proteins.

There are few reagents or assay kits available for quantitating thiols in biological systems. However, all the commercial kits either lack sensitivity or have tedious protocols. Our AmpliteTM Fluorimetric Thiol Quitation Kit provides an ultrasensitive fluorimetric assay for quantitating thiols that exist either in a small molecule or on a protein. The kit uses a proprietary non-fluorescent dye that becomes strongly fluorescent upon reacting with thiol. The kit provides a sensitive, one-step fluorimetric method to detect as little as 1 picomole of cysteine or GSH in a 100 µL assay volume (10 nM; Figure 1). The assay can be performed in a convenient 96-well or 384-well microtiter-plate format and easily adapted to automation with no separation steps required. Its signal can be easily read by fluorescence microplate reader with Ex/Em = 490 nm/520 nm.

Kit Key Features			
Broad Application:	Can be used for quantifying thiol and sulfide in a variety of biological systems (e.g., plasma, urine and cell extracts)		
Sensitive:	Detect as low as 1 picomole of thiol.		
Continuous:	Easily adapted to automation with no separation required.		
Convenient:	Formulated to have minimal hands-on time. No wash is required.		
Non-Radioactive:	No special requirements for waste treatment.		

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Kit Components

Components	Amount	
Component A: Thiolite Green TM	1 vial	
Component B: Assay Buffer	1 bottle (25 ml)	
Component C: GSH standard	1 vial (62 µg)	
Component D: DMSO	1 vial (100 μL)	

Assay Protocol for one 96-well plate

Brief Summary

Prepare Thiolite GreenTM reaction mixture (50 μ L) \rightarrow Add GSH standards or test samples (50 μ L) \rightarrow Incubate at room temperature for 10 min-1 h \rightarrow Read fluorescence at Ex 490 nm/Em 520nm nm

Note: Thaw all the kit components to room temperature before starting your experiment.

1. Prepare GSH standard stock solution:

<u>Prepare GSH standard stock solution</u>: Add 200 μ L of ddH2O into the GSH standard vial (Component C) to make 1 mM (1 nmol/ μ L) stock solution. *Note: The unused GSH solution should be divided as single use aliquots and stored at -20°C*.

2. Prepare 400X Thiolite Green[™] stock solution:

<u>Prepare Thiolite GreenTM stock solution</u>: Add 25 μ L of DMSO (Component D) into the Thiolite Green vial (Component A) to make 400X stock solution. *Note: The unused Thiolite Green solution should be divided as single use aliquots and stored at*

-20°C and avoid from light.

3. Prepare GSH reaction mixture:

<u>Prepare the GSH reaction mixture</u>: Add 12.5 μ L of 400X Thiolite GreenTM stock solution (from step 2) into 5 ml assay buffer (Component B), mixed well.

4. Prepare Serial GSH (0 to 30 $\mu M)$ solutions

4.1 Add 30 μ L of GSH standard stock solution (from step 1) to 970 μ l assay buffer (Component B) to generate 30 μ M (30 pmol/ μ L) standard.

Note: Diluted GSH standard solution is unstable, should be used within 4 hours.

- 4.2 Take 200 μL of 30 μM solution to perform 1:3 serial dilutions to get 10, 3, 1, 0.3, 0.1, 0.03, 0.01 and 0 standard GSH solutions.
- 4.3 Add GSH standards and GSH-containing or other thiol-containing test samples into a 96-well solid black microplate as described in Tables 1 and 2 *Note: Treat your cell or tissue samples as desired.*

BL	BL	TS	TS	 			
GS1	GS1			 			
GS2	GS2						
GS3	GS3						
GS4	GS4						
GS4 GS5	GS5						
GS6	GS6						
GS7	GS7						

Table 1. Layout of GSH standards and test samples in a solid black 96-well microplate:

Note: GS= GSH standards, BL=Blank control, TS=test samples.

Table 2. Reagent composition for each well:

GSH Standard	Blank Control	Test Sample
Serial dilutions* (50 µL)	Assay buffer: 50 µL	50 μL

*Note: Add the serially diluted GSH standards from 0.01 μ M to10 μ M into wells from NS1 to NS7 in duplicate.

5. Run GSH Assay

- 5.1 Add 50 μ L of GSH assay mixture (from step 3) to each well of the GSH standard, blank control, and test samples (see step 4.3) so that the total GSH assay volume is 100 μ L/well *Note: For a 384-well plate, add 25 \muL sample, 25 \muL of GSH reaction mixture per well.*
- 4.1 Incubate the reaction for 10 minutes to 1 hour at room temperature, protected from light.
- 4.2 Monitor the fluorescence increase with 490 nm excitation and 520 nm emission by using a fluorescence plate reader.

6. Run Data Analysis

The fluorescence in blank wells (with the assay buffer only) is used as a control, and is subtracted from the values for those wells with the GSH reactions. The typical data are shown in Figure 1 (GSH standard curve).

Note: The fluorescence background increases with time, thus it is important to subtract the fluorescence intensity value of the blank wells for each data point.

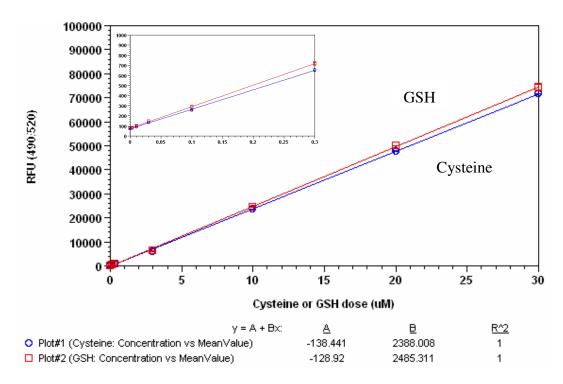


Figure 1. GSH and Cysteine dose response on 96-well black plate was measured with AmpliteTM Fluorimetric Thiol Quantitation Assay Kit using a BMG LabTech NOVOStar microplate reader. As low as 10 nM (1 pmol/well) of GSH or Cysteine can be detected with 10 minutes incubation time (n=3). The insert shows the low levels of thiol detection.

References:

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